Cruise: WS18120
Ship: R/V Walton Smith
Dates: April 30th – May 3rd, 2018
Expocode: 33WA20180430
Chief Scientist: Chris Kelble
Equipment: CTD and Flow Through (FT)
Total number of stations: 16
Location: Southwest Florida Gulf of Mexico coastal region

Samples were collected for Dr. Leticia Barbero for the Ocean Acidification Program during the South Florida Project (SFP) water quality cruises in the SW Gulf of Mexico lead by Dr. Chris Kelble.

Sample Collection

The discrete samples were collected from the CTD/rosette system onboard the R/V Walton smith by Chris Kelble. The date and time listed in the data file are UTC when each sample bottle was collected.

DIC:
16 locations, 16 samples each 500-ml, no duplicate samples.
Sample_ID#: 90101, etc.; Station, cast number and Niskin bottle number
PI: Dr. Rik Wanninkhof
Analyzed by: Charles Featherstone and Patrick Mears

pH:
16 locations, 16 samples each 500-ml, no duplicate samples.
Sample_ID#: 90101, etc.; Station, cast number and Niskin bottle number
PI: Dr. Rik Wanninkhof
Analyzed by: Charles Featherstone and Patrick Mears

TAkk:
16 locations, 16 samples each 500-ml, no duplicate samples.
Sample_ID#: 90101, etc.; Station, cast number and Niskin bottle number
PI: Dr. Rik Wanninkhof
Analyzed by: Dr. Leticia Barbero, Charles Featherstone and Patrick Mears

Sample Analysis
DIC:

<table>
<thead>
<tr>
<th>Instrument ID</th>
<th>Date</th>
<th>Certified CRM (µmol/kg)</th>
<th>CRM Value (µmol/kg)</th>
<th>CRM Offset (µmol/kg)</th>
<th>Blank (Counts)</th>
<th>Avg. Sample Analysis Time</th>
</tr>
</thead>
<tbody>
<tr>
<td>AOML 5</td>
<td>08/10/2018</td>
<td>2042.41</td>
<td>2043.27</td>
<td>0.86</td>
<td>20.0</td>
<td>8</td>
</tr>
<tr>
<td>AOML 6</td>
<td>08/10/2015</td>
<td>2042.41</td>
<td>2043.40</td>
<td>0.99</td>
<td>20.0</td>
<td>9</td>
</tr>
</tbody>
</table>
Analysis date: 08/10/2018
Coulometer used: DICE–CM5011- AOML 5
Blanks: 12.7, 20.0 counts/min
CRM # 665 was used and with an assigned value of (includes both DIC and salinity):
Batch 173, c: 2042.41 µmol/kg, S: 33.414
CRM values measured: AOML 5: offset 0.86 µmol/kg (2043.27 µmol/kg).
Average run time, minimum run time, maximum run time: 8, 7 and 10 min.

Analysis date: 08/10/2018
Coulometer used: DICE–CM5011- AOML 6
Blanks: 12.0, 20.0 counts/min
CRM # 723 was used and with an assigned value of (includes both DIC and salinity):
Batch 173, c: 2042.41 µmol/kg, S: 33.414
CRM values measured: AOML 6: offset 0.99 µmol/kg (2043.40 µmol/kg).
Average run time, minimum run time, maximum run time: 9, 7 and 17 min.

Reproducibility: (# samples and average difference): No duplicate samples were collected.

CRM, salinity and HgCl2 correction applied: Salinity correction was applied using TSG salinity.

Remarks

The volume correction was applied due to added HgCl2 (Measured DIC*1.00037). The first CRM of each cell was used for a CRM correction.

The DIC instruments were stable: the gas loop and CRM values did not change significantly throughout the life span of each cell.

The blank on AOML 5 (08/10/2018) was raised from 12.7 to 20.0 before running the CRM.

The blank on AOML 6 (08/10/2018) was raised from 12.0 to 20.0 before running the CRM.

pH:

Analysis date: 08/10/2018
A CRM was analyzed before sample analysis.
CRM #536, Batch 173 pH = 7.8787

Spectrophotometer used: HP Agilent 8453
Salinity and temperature of pH samples analyzed.

<table>
<thead>
<tr>
<th>Sample ID</th>
<th>Station</th>
<th>Bottle #</th>
<th>Salinity</th>
<th>Temperature</th>
</tr>
</thead>
<tbody>
<tr>
<td>CRM-173-536</td>
<td>CRM</td>
<td>536</td>
<td>33.414</td>
<td>20.038</td>
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<tr>
<td>54</td>
<td>54</td>
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<td>32.773</td>
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<tr>
<td>55</td>
<td>55</td>
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<td>34.468</td>
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<td>56</td>
<td>11</td>
<td>35.84</td>
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<tr>
<td>57</td>
<td>57</td>
<td>12</td>
<td>36.343</td>
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<td>13</td>
<td>36.453</td>
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<td>37.06</td>
<td>20.044</td>
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<td>20.034</td>
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<td>57.3</td>
<td>14</td>
<td>36.265</td>
<td>20.044</td>
</tr>
</tbody>
</table>

**Reproducibility:** (# samples and average difference): No duplicates were collected.

**Remarks**

The equations of Liu et al, 2011 formulated using the purified m-cresol purple indicator was used to determine pH of the samples. pH samples were analyzed at 20°C at Full Scale (pH 0-14). The pH was reported at 25°C.

Temperature for each sample was measured before analysis using a Hart Scientific Fluke 1523 reference thermometer.

Approximately 80 mL of sample was extracted from each DIC sample bottle by automatic syringe before DIC analysis to determine the pH.

**Talk:**
Analysis date: 08/17/2018
Titration system used: Open cell
CRM #41 Batch 153, Salinity = 33.357, cert. TA = 2225.59μmol/kg.
CRM #739 Batch 150, Salinity = 33.343, cert. TA = 2214.71μmol/kg

On 08/17/2018 CRM #41, Batch 153 was analyzed before the samples and CRM #739,
Batch 150 was run at the end of analysis on system 1. The TA for the water samples was corrected using the daily averaged ratios between the certified and measured values of the CRMs run on system 1 cell. The following table shows the CRM measurements for each day and cell.

| Cell | System | Date       | Time       | Bottle # | TA    | |ΔCRM| |
|------|--------|------------|------------|----------|-------|------|------|
| 1    | 08/17/2018 | 09:28:20  | 41         | 2229.30  | 3.71  |      |      |
| 1    | 08/17/2018 | 19:57:15  | 739        | 2215.77  | 1.06  |      |      |

**Reproducibility:** No duplicates were collected.

**Remarks**

**Comments**

The latitude, longitude, date, and time reported with the DIC, pH and TA pots measurements were taken from the sample field log. The field log values are provided for reference; no post-cruise assurance of accuracy has been done to this data. The Niskin bottles are approximately one half meter above the CTD sensors on the rosette. Therefore, Temp and Sal are bin-averaged CTD values representing the next shallower depth from that recorded by the CTD (CTD Depth) at the time the Niskin bottles were fired with the exception of the surface values, which are the same as the CTD Depth values (as per the log sheet). The FT sample salinity and temperature were collected from the ship’s TSG.

The Sample ID is the sample station, cast number and Niskin bottle number for the discrete samples.

Corresponding UW pCO2 data can be found at the following website http://www.aoml.noaa.gov/ocd/ocdweb/occ.html

**Nutrients:**

**Analysis Date:** May 22\(^{nd}\), 2018

Nutrient samples were analyzed using a Seal Analytical high resolution digital colorimeter auto-analyzer 3 (AA3). A series of standards for each method were run before sample analysis to obtain a calibration curve for data reduction. Method 353.4 was used to determine the concentration of nitrate and nitrite for each station (Zhang et al., 1997b). This method used automated, gas-segmented, continuous flow colorimetry for the analysis of nitrate and nitrite. Samples were first passed through a copper-coated cadmium reduction column. Nitrate was reduced to nitrite in a buffer solution. The nitrite was then determined by diazotizing with sulfanilamide and coupling with N-1-naphthylethlenediamine dihydrochloride to form a color azo dye. The absorbance measured at 550 nm is linearly proportional to the concentration of nitrite +
Nitrate in the sample. Nitrate concentrations are obtained by subtracting nitrite values, which have been separately determined without the cadmium reduction procedure, from the nitrite + nitrate values.

Method 365.5 was used to determine the concentration of orthophosphate for each station (Zimmermann and Keefe, 1997; Zhang et al., 2001). This method used automated colorimetric and continuous flow analysis for the determination of low-level orthophosphate concentrations. Ammonium molybdate and antimony potassium tartrate react in an acidic medium with orthophosphate to form an antimony-phospho-molybdate complex. This complex was reduced to a blue-colored complex by ascorbic acid. The absorbance measured at 880 nm is proportional to the phosphate concentration in the sample.

Method 366.0 was used to determine the concentration of soluble silica for each station (Zhang and Berberian, 1997). This method used automated, gas-segmented, continuous flow colorimetry for the analysis of dissolved silicate concentration. In this method, $\beta$-molybdosilicic acid was formed by the reaction of the silicate contained in the sample with molybdate in acidic solution. The $\beta$-molybdosilicic acid was then reduced by ascorbic acid to form molybdenum blue. The absorbance of the molybdenum blue, measured at 550 nm, is linearly proportional to the concentration of silicate in the sample.


UPDATE:
Between March and June of 2021, all of the data for the discrete samples was put into a uniform format. The supporting information was checked for accuracy, especially the expocode, date, time, and positions. Additionally, pH results were recalculated to 20 and 25 degrees Celsius.